

# A dual role of 5-hydroxytryptamine receptor 3 in serotonin induced ion transport in rat distal colon

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Received 18 September 2007; received in revised form 23 December 2007; accepted 21 January 2008

Available online 5 February 2008

## Abstract

5-hydroxytryptamine (5-HT)-evoked intestinal secretion can be divided into neural and non-neural pathway. Recently, 5-HT<sub>3</sub> receptor in neural pathway received much attention as a possible target in bowel diseases. The present study aims to investigate the effects of 5-HT<sub>3</sub> receptor in different enteric neural plexus (myenteric plexus and submucosal plexus) on rat colonic ion transport by using rat intact colon and mucosa/submucosa preparations. Ussing chamber and real-time PCR techniques were performed in our present study. Surprisingly, we found that in mucosa/submucosa preparations, 5-HT-induced  $\Delta I_{SC}$  (change in short-circuit current) was not inhibited, but further increased by pretreatment with 5-HT<sub>3</sub> receptor antagonists, MDL72222 and Tropanyl-3, 5-dimethylbenzoate. And this response was significantly attenuated in the presence of tetrodotoxin (TTX). Conversely, in rat intact colon, 5-HT<sub>3</sub> receptor antagonists significantly inhibited 5-HT-induced  $\Delta I_{SC}$ . The results from real-time PCR proved the existence of 5-HT<sub>3</sub> receptor in muscularis externa and submucosa. Taken together, 5-HT<sub>3</sub> receptors possess a role of dual regulation on electrolyte secretion in rat distal colon, the neural stimulatory effect of 5-HT<sub>3</sub> receptor in myenteric plexus and the inhibitory effect of 5-HT<sub>3</sub> receptor in submucosal plexus.

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**Keywords:** 5-HT<sub>3</sub> receptor; Ion transport; Neural pathway; Inhibitory; Ussing chamber

## 1. Introduction

5-HT (5-hydroxytryptamine, serotonin) is an important active enteric amine in the gastrointestinal system. It has been suggested to play a role in the regulation of colonic motor and secretory functions in several species and humans. In mammals, about 60–90% (up to 10 mg in man) of the total amount of 5-HT in the body

is in gastrointestinal tract and most of it exists in enterochromaffin (EC) cells (Kellum et al., 1999). 5-HT is secreted from EC cells in response to variety of luminal mechanical and chemical stimulations within the gastrointestinal wall. It is also related to several gastrointestinal dysfunctions such as emesis, motility disorders, diarrhea, and more recently, irritable bowel syndrome (Hoyer et al., 2002). Other major sites of 5-HT are the central nervous system, myenteric nerve system, platelets and so on.

5-HT produces its effects through membrane-bound receptors in different organs. In vitro studies, 5-HT-evoked intestinal secretion was divided into two pathways: neural and non-neural pathway (Budhoo et al., 1996a; Hansen and Skadhauge, 1997; Kiso et al., 1997). The non-neural pathway-mediated stimulatory effect in our previous study was found to be mainly via 5-HT<sub>4</sub> receptor residing at the level of the colonocyte (Ning et al., 2004). while the neural pathway was chiefly mediated by 5-HT<sub>3</sub> receptor (Budhoo et al., 1996a; Kiso et al., 1997). Using intact

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rat distal colon, Budhoo et al. found that specific 5-HT<sub>3</sub> receptor agonist, 2-methyl-5-HT produced a concentration-dependent change in  $I_{SC}$  and its action was antagonized by tetrodotoxin (TTX, an inhibitor of neural conduction) (Budhoo et al., 1996a; Kiso et al., 1997), which indicates that the 5-HT<sub>3</sub> receptor produces a stimulatory effect in neural pathway.

Recently, the neural 5-HT<sub>3</sub> receptor received much attention as a possible target in bowel diseases (Michel et al., 2005). And 5-HT<sub>3</sub> receptor antagonists have been used to relieve symptoms associated with diarrhea-predominant irritable bowel syndrome (Hicks et al., 2002). But the exact mechanism underlying 5-HT<sub>3</sub> receptor neurally mediated ion transports in colon is still unclear.

The enteric nervous system (ENS) is the part of the nervous system that directly controls the gastrointestinal system. The neurons of the ENS are collected into two types of ganglia: myenteric (Auerbach's) and submucosal (Meissner's) plexuses. Myenteric plexuses are located between the inner and outer layers of the muscularis externa, while submucosal plexuses are located in the submucosa (Crone et al., 2003). The present study aims to investigate the effects of 5-HT<sub>3</sub> receptors in different enteric neural plexus (myenteric plexus and submucosal plexus) on rat colonic ion transport by using intact colon and mucosa/submucosa preparations.

## 2. Materials and methods

### 2.1. Animals

Animal protocols followed guidelines established by the NIH and were approved by Animal Care and Use Committee, Capital Medical University. Adult male Sprague–Dawley rats (Laboratory Animal Services Center, Capital Medical University) ranging in weight from 200 to 300 g had free access to standard rodent laboratory food and water until the day of the experiment. The animals were killed by cervical dislocation. The distal colon was removed and defined as the ~7 cm-long segment proximal to the lymph node (typically situated 3 cm apart from the anus). Then the distal colon was divided into four segments, termed DC<sub>1</sub> (adjacent to the lymph node), DC<sub>2</sub>, DC<sub>3</sub> and DC<sub>4</sub>, respectively. Preliminary results indicated that the differences existed in the four segments (Yang et al., 2006). But the responses of DC<sub>3</sub> and DC<sub>4</sub> to 5-HT were similar and stable. Therefore, in the present study rat DC<sub>3</sub> and DC<sub>4</sub> were taken to investigate the roles of 5-HT<sub>3</sub> receptors. Every DC<sub>3</sub> and DC<sub>4</sub> was cut along the mesenteric border into a flat sheet and flushed with ice-cold Krebs–Henseleit solution (K–HS) containing (in mmol/l): 117 NaCl, 4.7 KCl, 1.2 MgCl<sub>2</sub>·6H<sub>2</sub>O, 1.2 NaH<sub>2</sub>PO<sub>4</sub>, 25 NaHCO<sub>3</sub>, 2.5 CaCl<sub>2</sub>·2H<sub>2</sub>O, 11.1 D-glucose. The tissue was pinned flat with the mucosal side down in a Sylgard-lined petri dish containing ice-cold oxygenated reperfusion.

### 2.2. Reagents

5-hydroxytryptamine (5-HT), indomethacin, MDL-72222 (Tropanyl 3,5-dichlorobenzoate), GR113808 ([1-[2-(methylsulfonyl-amino)ethyl]-4-piperidinyl] methyl 1-methylindole-3-carboxylate) and tetrodotoxin (TTX) were obtained from Sigma Chemical

Company (St. Louis, MO, USA). Tropanyl-3, 5-dimethylbenzoate was purchased from Tocris Cookson Inc. (Ellisville, Missouri, USA). Stock solutions of some chemicals (indomethacin, MDL-72222, GR113808, Tropanyl-3, 5-dimethylbenzoate) were dissolved in dimethyl sulfoxide (DMSO). Final DMSO concentrations never exceeded 0.1% (vol/vol). Preliminary experiments indicated that the vehicle did not alter any baseline electrophysiological parameters.

### 2.3. Ussing chamber experiments

Flat sheets of mucosa/submucosa preparation or intact colon were mounted in modified Ussing chambers with a cross-sectional area being 0.5 cm<sup>2</sup>. The mucosal and serosal surfaces of tissue were bathed with 5 ml K–HS recirculated from a reservoir maintained at 37 °C and bubbled with 95% O<sub>2</sub> and 5% CO<sub>2</sub> to maintain the pH of the solution at 7.4. Drugs were added directly to the apical or basolateral side of epithelium. Responses were recorded continuously. Transepithelial potential difference for every colonic mucosa was measured with the Ag/AgCl reference electrodes (Physiologic Instruments, P2020S) connected to a preamplifier that was in turn connected to a voltage-clamp amplifier VCC MC6 (Physiologic Instruments, San Diego, CA, USA). The change in short-circuit current ( $\Delta I_{SC}$ ) was calculated as difference between before and after stimulation.  $I_{SC}$  was normalized as current per unit area of epithelial ( $\mu A/cm^2$ ), which allowed the curve area for 15 min to be calculated ( $\mu A \min$ ).

### 2.4. RNA extraction and preparation of cDNA

Two preparations from 9 rats were collected in phosphate buffer solution (PBS), which had been treated with 0.1% diethyl pyrocarbonate (DEPC-PBS), namely the intact colon and mucosa/submucosa preparations. Preparations were cut open along the mesenteric border, and the contents were flushed out with DEPC-PBS and immediately snap frozen in liquid nitrogen. RNA from gut tissue was harvested using the Trizol RNA purification system (Invitrogen, Carlsbad, CA, USA) according to the manufacturer's instructions.

### 2.5. Real-time polymerase chain reaction

Real-time PCR was used to quantify mRNA encoding 5-HT<sub>3</sub> receptors in the intact and mucosa/submucosa preparations of rat distal colon. The expression of 5-HT<sub>3</sub> was normalized to that of  $\beta$ -actin, a housekeeping gene that is not thought to be subject to regulation. Transcripts encoding 5-HT<sub>3</sub> receptors in samples of rat colon were comparatively quantified by real-time PCR with the Brilliant SYBR Green QPCR Master Mix kit (Stratagene, La Jolla, CA, USA) using a Light Cycler instrument (Stratagene).

Amplifications were performed in a final volume of 20  $\mu l$  of a commercial reaction mixture according to the manufacturer's instructions. The primers for the amplification of cDNA encoding  $\beta$ -actin and 5-HT<sub>3</sub> receptors were used at a final concentration of 0.2  $\mu mol/l$ . 0.25  $\mu l$  of the cDNA prepared from tissue was added to the mixture. Data were analyzed with computer assistance using the MxPro QPCR software (version 3.0, Mx3000P system,

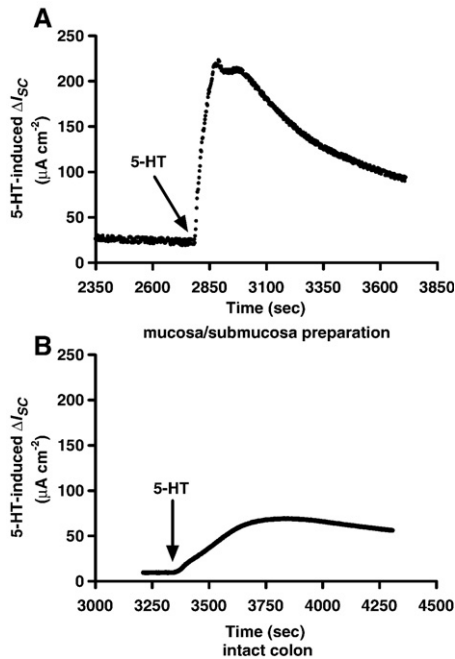


Fig. 1. 5-HT-induced  $I_{sc}$  responses in rat mucosa/submucosa preparations (A) and intact colon (B). Even at 100  $\mu mol/l$ , 5-HT-induced  $I_{sc}$  responses in rat intact colon were smaller than that (10  $\mu mol/l$ ) in rat mucosa/submucosa preparation.

Stratagene). Primer sequences were:  $\beta$ -actin forward primer: 5'-TTC AAC ACC CCA GCC ATG T-3', reverse primer: 5'-GTG GTA CGA CCA GAG GCA TAC A-3'; 5-HT<sub>3</sub> receptor forward

primer 5'-TGC ATA CCA TCC AGG ACA TCA-3', reverse primer: 5'-CTC TTG TCC GAC CTC ACT TCT TC-3'.

## 2.6. Statistical analysis

The data were expressed as means  $\pm$  standard error of mean (S.E.M.). "n" refers to the number of tissue preparations. Comparisons between groups of data were made via Student's paired or unpaired *t*-test. *P*-values  $<0.05$  were considered statistically significant.

## 3. Results

### 3.1. 5-HT-induced $I_{sc}$ responses in rat intact colon and mucosa/submucosa preparations

To investigate the role of 5-HT<sub>3</sub> receptor in regulation of ion transport, rat intact colon and mucosa/submucosa preparations were used respectively. In both of the preparations, indomethacin (10  $\mu mol/l$ ), a cyclooxygenase (COX) inhibitor, was routinely added to the basolateral side to abolish the effects of endogenous prostaglandins. 5-HT and antagonists of 5-HT receptors were added to the basolateral sides of the tissues in the present study since basolateral addition, but not apical application, of 5-HT was able to elicit an increase in  $I_{sc}$ . (Ning et al., 2004) After equilibration for 30 min, 5-HT (10  $\mu mol/l$ )-produced  $I_{sc}$  increase was  $1240.0 \pm 91.3 \mu A min$  ( $n=12$ ) in rat mucosa/submucosa preparations. However, in intact colon, 5-HT, in 10  $\mu mol/l$ , -induced  $I_{sc}$  response was very small and can be ignored (data not shown), and 5-HT, in

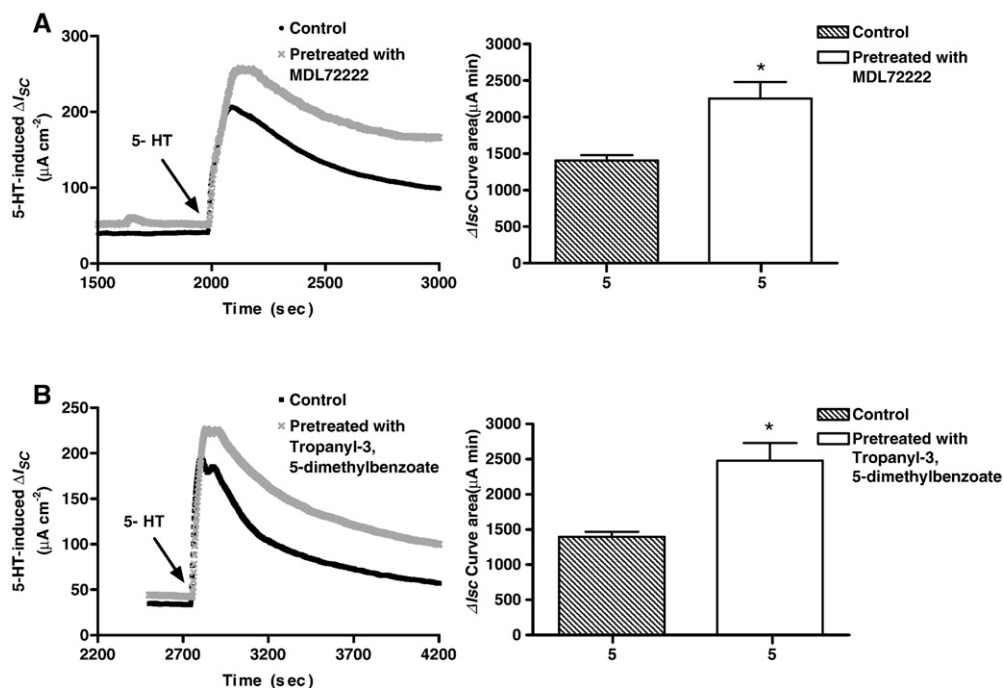


Fig. 2. Effect of 5-HT<sub>3</sub> receptor antagonists on 5-HT-induced  $I_{sc}$  responses in rat distal colon mucosa/submucosa preparations. (left) Representative  $I_{sc}$  recording with arrows indicating the time for the basolateral application of 5-HT (10  $\mu mol/l$ ) to rat mucosa/submucosa preparations basolateral pretreated with MDL72222 (10  $\mu mol/l$ , A) or Tropanyl-3, 5-dimethylbenzoate (10  $\mu mol/l$ , B), respectively. (right) Summary of the effects of MDL72222 (10  $\mu mol/l$ , A) and Tropanyl-3, 5-dimethylbenzoate (10  $\mu mol/l$ , B) on 5-HT-induced  $\Delta I_{sc}$ . Values are means  $\pm$  S.E.M.; \*,  $P < 0.05$ .

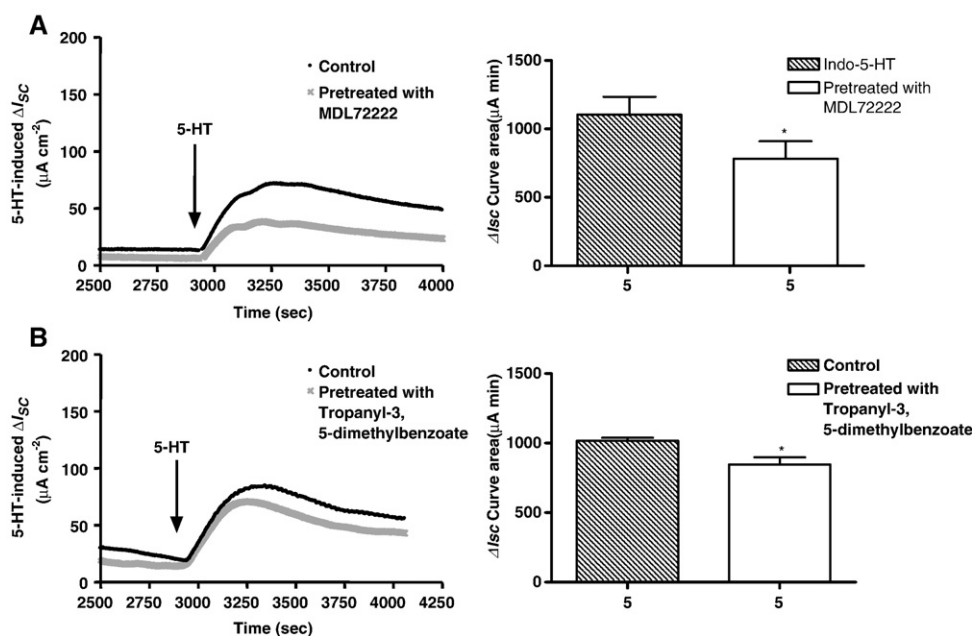


Fig. 3. Effect of 5-HT<sub>3</sub> receptor antagonists on 5-HT-induced  $I_{SC}$  responses in rat intact colon preparations. (left) Representative  $I_{SC}$  recording with arrows indicating the time for the basolateral application of 5-HT (10  $\mu$ mol/l) to rat intact colon preparations basolateral pretreated with MDL72222 (10  $\mu$ mol/l, A) or Tropanyl-3, 5-dimethylbenzoate (10  $\mu$ mol/l, B), respectively. (right) Summary of the effects of MDL72222 (10  $\mu$ mol/l, A) and Tropanyl-3, 5-dimethylbenzoate (10  $\mu$ mol/l, B) on 5-HT-induced  $\Delta I_{SC}$ . Values are means  $\pm$  S.E.M.; \*,  $P < 0.05$ .

100  $\mu$ mol/l, induced change in  $I_{SC}$  was  $997.0 \pm 83.5$   $\mu$ A min ( $n = 9$ ). (Fig. 1)

### 3.2. Effects of 5-HT<sub>3</sub> receptor antagonists on 5-HT-induced $I_{SC}$ responses in rat intact colon and mucosa/submucosa preparations

The pharmacological profile of 5-HT<sub>3</sub> receptor in rat intact colon and mucosa/submucosa preparation was evaluated

by using specific antagonists (MDL72222 and Tropanyl-3, 5-dimethylbenzoate) to 5-HT<sub>3</sub> receptor.

In rat mucosa/submucosa preparations, pretreatment with 5-HT<sub>3</sub> receptor antagonists MDL72222 (10  $\mu$ mol/l) and Tropanyl-3, 5-dimethylbenzoate (10  $\mu$ mol/l) for 5 min did not inhibit, but increased 5-HT (10  $\mu$ mol/l)-induced  $\Delta I_{SC}$  from  $1406.0 \pm 72.7$   $\mu$ A min and  $1395.0 \pm 73.7$   $\mu$ A min to  $2255.0 \pm 227.0$   $\mu$ A min ( $n = 5$ ,  $P < 0.05$ , Fig. 2A) and  $2479.0 \pm 249.5$   $\mu$ A min ( $n = 5$ ,

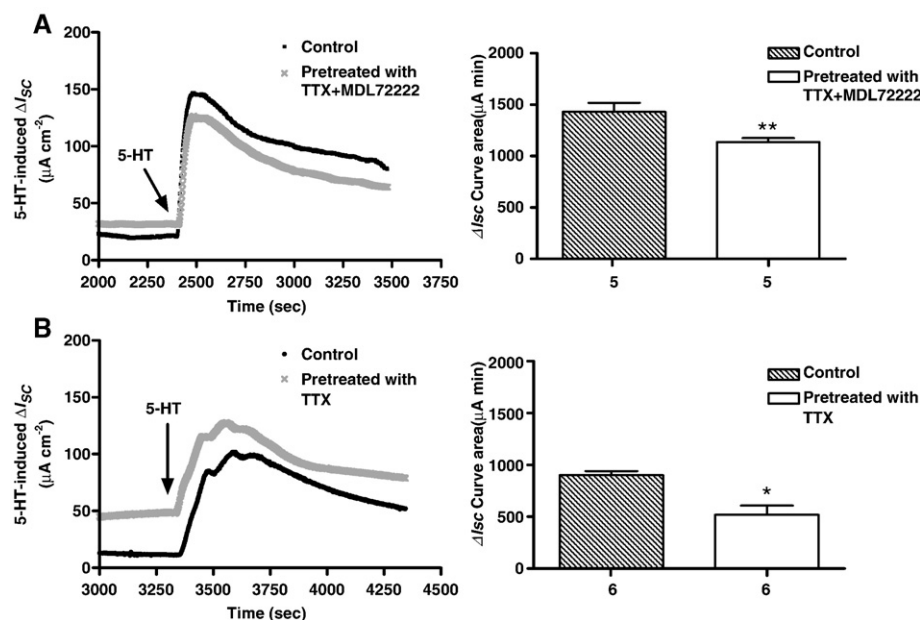


Fig. 4. Role of neural pathway in 5-HT-induced  $I_{SC}$  responses. (left) Representative  $I_{SC}$  recording with arrows indicating the time for the basolateral application 5-HT (10  $\mu$ mol/l) to rat mucosa/submucosa preparations basolateral pretreated with TTX (1  $\mu$ mol/l) and MDL72222 (10  $\mu$ mol/l, A) or rat intact colon preparations basolateral pretreated with TTX (1  $\mu$ mol/l, B), respectively. (right) Summary of the effects of TTX (1  $\mu$ mol/l) and MDL72222 (10  $\mu$ mol/l, A) or TTX (1  $\mu$ mol/l, B) on 5-HT-induced  $\Delta I_{SC}$  in rat mucosa/submucosa or intact colon preparations, respectively. Values are means  $\pm$  S.E.M.; \*,  $P < 0.05$ ; \*\*,  $P < 0.01$ .



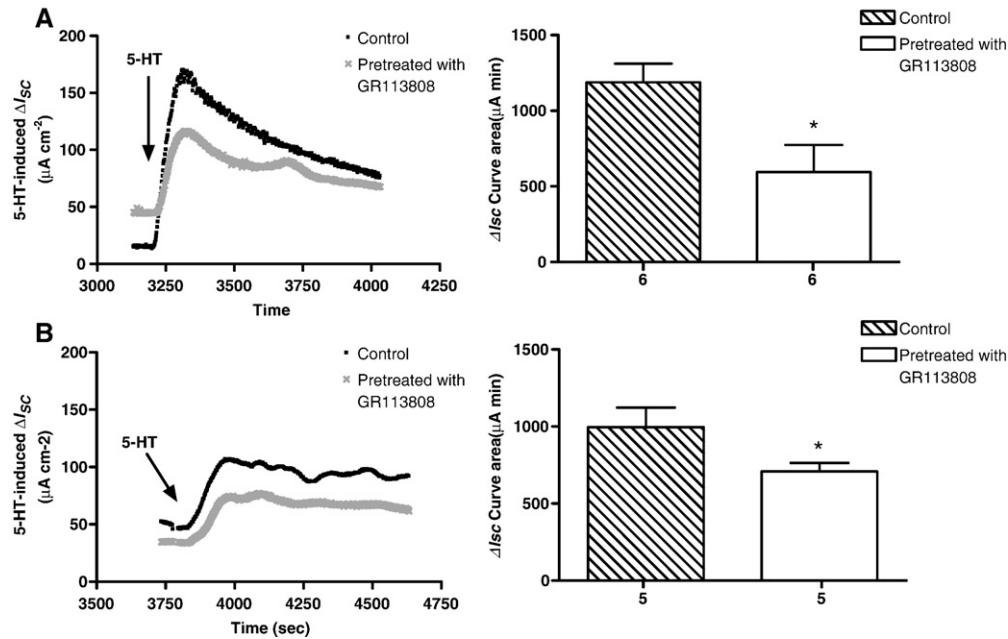


Fig. 5. Effect of 5-HT<sub>4</sub> receptor antagonist on 5-HT-induced  $I_{SC}$  responses in rat mucosa/submucosa and intact colon preparations. (left) Representative  $I_{SC}$  recording with arrows indicating the time for the basolateral application of 5-HT to rat mucosa/submucosa (A) and intact colon (B) preparations basolateral pretreated with GR 113808 (0.1  $\mu mol/l$ ), respectively. (right) Summary of the effects of GR 113808 (0.1  $\mu mol/l$ ) on 5-HT-induced  $\Delta I_{SC}$  in rat mucosa/submucosa (A) and intact colon (B) preparations. Values are means  $\pm$  S.E.M.; \*,  $P < 0.05$ .

$P < 0.05$ , Fig. 2B), respectively. Whereas, in rat intact colon, both of the 5-HT<sub>3</sub> receptor antagonist MDL72222 (10  $\mu mol/l$ ) and Tropanyl-3, 5-dimethylbenzoate (10  $\mu mol/l$ ) significantly decreased 5-HT (100  $\mu mol/l$ )-induced  $I_{SC}$  from  $1105.0 \pm 130.0 \mu A min$  and  $1017.0 \pm 21.8 \mu A min$  to  $782.5 \pm 128.0 \mu A min$  ( $n = 5$ ,  $P < 0.05$ , Fig. 3A) and  $846.7 \pm 50.4 \mu A min$  ( $n = 5$ ,  $P < 0.05$ , Fig. 3B), respectively.

### 3.3. Effects of neural pathways on 5-HT-induced $I_{SC}$ responses in rat intact colon and mucosa/submucosa preparations

It has been reported that 5-HT-induced  $I_{SC}$  responses is mainly neurally mediated by the 5-HT<sub>3</sub> receptor (Budhoo et al., 1996a). To study the role of enteric neural pathway, tetrodotoxin (TTX, 1  $\mu mol/l$ ), an inhibitor of neural conduction, was chosen. In rat mucosa/submucosa preparations basolateral pretreated with TTX, the increased effect of 5-HT<sub>3</sub> antagonist on 5-HT-induced responses was absent. On the contrary, the inhibitory effect was observed, 5-HT (10  $\mu mol/l$ )-induced  $I_{SC}$  was significantly reduced by MDL72222 (10  $\mu mol/l$ , basolateral side) from  $1429.0 \pm 89.7 \mu A min$  to  $1135.0 \pm 41.3 \mu A min$  ( $n = 5$ ,  $P < 0.01$ , Fig. 4A). In rat intact colon, pretreated with TTX, 5-HT (100  $\mu mol/l$ )-induced current was decreased from  $900.9 \pm 39.5 \mu A min$  to  $518.4 \pm 90.6 \mu A min$  ( $n = 6$ ,  $P < 0.05$ , Fig. 4B).

### 3.4. Effects of 5-HT<sub>4</sub> receptor antagonists in rat intact colon and mucosa/submucosa preparations

Pretreated with 5-HT<sub>4</sub> antagonist GR113808 (0.1  $\mu mol/l$ , basolateral side), 5-HT (10  $\mu mol/L$ )-induced  $\Delta I_{SC}$  was significantly inhibited by about 50% (from  $1188.0 \pm 122.5 \mu A min$  to

$595.2 \pm 178.6 \mu A min$ ,  $n = 6$ ,  $P < 0.05$ , Fig. 5A) in rat mucosa/submucosa preparations and 28.84% (from  $995.2 \pm 127.9 \mu A min$  to  $707.6 \pm 55.3 \mu A min$ ,  $n = 5$ ,  $P < 0.05$ , Fig. 5B) in rat intact colon.

### 3.5. The mRNA expression of 5-HT<sub>3</sub> receptor in different preparations of rat colon

In rat gastrointestinal tract, 5-HT<sub>3</sub> receptors were found in numerous myenteric and submucosal neurons and were abundant in fibers within the myenteric plexus (Thompson and Lummis, 2006). It has been reported that the 5-HT<sub>3</sub> receptor subunits may be the same (homopentameric 5-HT<sub>3a</sub> receptors) or different (heteropentameric receptors, usually comprising of 5-HT<sub>3a</sub> and 5-HT<sub>3b</sub> receptor subunits), with the latter having a number of distinct properties (Siriwardena et al., 1993). Therefore, we chose the 5-HT<sub>3a</sub> receptor subunit as the marker of the 5-HT<sub>3</sub> receptor. Real-time PCR was used to compare the rat distal colonic mRNA expressions of 5-HT<sub>3</sub> receptor in intact colon, mucosa/submucosa and mucosa preparations in the present study. The relative mRNA quantities of 5-HT<sub>3a</sub> receptor subunit were normalized to the  $\beta$ -actin in each preparation. It is obvious that the mRNA expression of 5-HT<sub>3a</sub> receptor subunit exists in each layer of rat distal colon. The rank order expression of 5-HT<sub>3a</sub> receptor subunit was intact colon > mucosa/submucosa preparations (M + S) > mucosa preparations (M).

## 4. Discussion

Generally, the Ussing chamber voltage-clamp model is demonstrated as a measure of electrolyte transport. The secretory response of anion is reflected by an increase in short-circuit current

( $I_{SC}$ ). Then, in the present study, Ussing chamber was used to measure the secretory responses induced by 5-HT and the effects of 5-HT receptor antagonists. The classic study by Budhoo et al. demonstrated that 5-HT-stimulated mucosal electrolyte transport is mediated by a neural and non-neural mechanism (1996a). The investigators also found that in intact colon the stimulatory effect of 5-HT on gastrointestinal tract mediated by neural pathway is regulated via the 5-HT<sub>3</sub> receptor (Siriwardena et al., 1991, 1993). In our present study, a similar stimulatory role of 5-HT<sub>3</sub> receptor was found in rat intact colonic preparation for 5-HT<sub>3</sub> receptor antagonists inhibited 5-HT-induced  $\Delta I_{SC}$ . Moreover, after incubation with a neuronal Na<sup>+</sup> channel blocker, TTX, 5-HT-induced  $\Delta I_{SC}$  in rat intact colon was slightly diminished. Arcuni et al. reported that TTX was able to abolish 5-HT<sub>3</sub> receptor agonist, 2-methyl-5-HT, induced changes in  $I_{SC}$  due to its effects of chemically blocking an intact intrinsic innervation (2000). These results suggest that the neural stimulatory effect of 5-HT probably comes from the activated 5-HT<sub>3</sub> receptors in enteric plexus. However, one striking finding in rat mucosa/submucosa preparations was the inhibitory effect of 5-HT<sub>3</sub> receptor on rat colonic secretion. Results from the present study show that 5-HT<sub>3</sub> receptor antagonists, MDL72222 and Tropicamide, increased, but not decreased 5-HT-induced  $\Delta I_{SC}$  in rat mucosa/submucosa preparations and the effects were significantly attenuated in the presence of TTX. It illustrates a possible mechanism of inhibitory effect on anion secretion depending on the submucosal plexus mediated by 5-HT<sub>3</sub> receptor. The inhibitory effect of 5-HT<sub>3</sub> receptor on 5-HT-induced secretion in rat distal colon has never been reported although Ishizawa reported an inhibitory action of 5-HT on spontaneous propulsive activities depending on an inhibitory neurotransmitter-mediated endogenous prostaglandins release from the circular muscle cells (1996).

Our results from real-time PCR provide the evidences for the mRNA expressions of 5-HT<sub>3a</sub> receptors in both intact colon and mucosa/submucosa preparations. The rank order of 5-HT<sub>3a</sub> receptor subunit expression, intact colon > mucosa/submucosa preparation (M+S) > mucosa preparation (M), suggested that the 5-HT<sub>3a</sub> receptor mRNA existed in both muscularis externa and submucosa. It has been demonstrated that 5-HT<sub>3</sub> receptor-like immunoreactivity occurs in enteric neurons of all layers in the rat colon (Mazzia et al., 2003). Michel et al. also found the positive staining for 5-HT<sub>3a</sub> and 5-HT<sub>3b</sub> receptor in all neurons of the human submucous plexus (2005). These results provide possibility to our hypothesis got from Ussing chamber experiments, in neural pathway the stimulatory effect of 5-HT on electrolyte secretion owing to the activated 5-HT<sub>3</sub> receptor in myenteric plexus and the inhibitory effect of 5-HT being mediated by 5-HT<sub>3</sub> receptor in the submucosal plexus.

On the other hand, previous studies from our laboratory have shown that 5-HT elicited an ion transport by acting directly on the colonic mucosa via 5-HT<sub>4</sub> receptors (Ning et al., 2004). In the present study, we further investigated whether or not the 5-HT<sub>4</sub> receptor could also mediate a stimulatory  $I_{SC}$  response with the mucosa/submucosa preparations. Expectably, the effects of 5-HT<sub>4</sub> receptor antagonist on 5-HT-induced ion transport in intact colon and mucosa/submucosa preparations were similar to that in colonic mucosa. It is consistent with the

reports that 5-HT-induced non-neural ion transport is mediated by 5-HT<sub>4</sub> receptor in rat distal colon and human jejunal mucosa (Budhoo et al., 1996b; Ning et al., 2004).

Summing up, 5-HT<sub>3</sub> receptors possess a role of dual regulation on electrolyte secretion in rat distal colon, the neural stimulatory effect of 5-HT<sub>3</sub> receptor in myenteric plexus and the inhibitory effect of 5-HT<sub>3</sub> receptor in submucosal plexus. This leads to an increase in our understanding of the role of 5-HT in normal gastrointestinal (GI) physiology. It is well known that 5-HT<sub>3</sub> and 5-HT<sub>4</sub> receptors play important roles in regulation of 5-HT on enteric water and electrolyte flux (Budhoo et al., 1996a). Up to now, both 5-HT agonists and antagonists have proven clinical efficacy to irritable bowel syndrome (irritable bowel syndrome with diarrhea, irritable bowel syndrome with constipation), though in both cases, the large number needed to treat suggests that improved ways are needed to target and characterize patients who are likely to respond (Spiller, 2007). Our amazing finding that 5-HT<sub>3</sub> receptors play dual roles in regulation of gastrointestinal secretion may be particularly significant in relation to the pathogenesis of various gastrointestinal dysfunctions and provide the clue for diagnosis and treatment to different kind of patients (Tonini and Pace, 2006).

## Acknowledgments

This work was supported by the National Natural Science Foundation of China (30570672 and 30770799), Beijing Natural Science Foundation Program (7063070), Scientific Research Common Program of Beijing Municipal Commission of Education (KM200610025001), the Ph.D. Programs Foundation of Ministry of Education of China (20050025001), Beijing Municipal Project for Developing Advanced Human Resources for Higher Education and National Basic Research Program of China-973 project (2006CB500700).

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